

1. Yeast Dextrose Chalk (YDC) agar medium:

WHAT YOU NEED FOR 1 LITRE	
Agar	15 g
Yeast Extract	10 g
CaCO ₃ (light powder)	20 g
D-Glucose (Dextrose)	20 g

1. Weigh out all ingredients into a suitable oversize container
2. Add 750 ml distilled or bottled water. Bring to boil to dissolve. (CaCO₃ will not completely dissolve).
3. Make up to 1 litre volume with water.
4. Dispense into bottles (300 ml or 500 ml medical flats supplied), with constant swirling to ensure even distribution of CaCO₃. Fill bottle half to two thirds full.
5. Autoclave at 121°C, 115 psi for 15 minutes.
6. Allow medium to cool to approximately 50°C
7. Swirl to ensure even distribution of CaCO₃ and avoid air bubbles. Pour into sterile petri plates (22 ml per 9.0 cm plate).
8. Leave plates to set in a laminar flow cabinet or similar, before use.
9. Store prepared plates inverted in polythene bags at room temperature. Prepared plates can be stored for several months provided they do not dry out.

2. MXP medium:

To prepare medium for *Xanthomonas Campestris pr phaseoli* (MXP) see the instructions in the article provided:

Claffin, L.E., Vidavar, A.K. and Sasser, M. (1987).

MXP, a semi-selective medium for *Xanthomonas campestris* pv. Phaseoli. *Phytopathology* 77:730-734

3. Cornmeal Agar and other media supplied:

Product information from Oxoid can be found on the Trust's website at www.kirkhoustrust.org, click on 'Protocols'.