DNA markers (SCARS) linked with disease resistance traits in bean (*Phaseolus vulgaris*) Updated: 1/20/05 by Phil Miklas

SCAR Name	Marker of Origin	Pathogen	Size (bp) / orientation	Sequences of SCARS	Tagged Locus	LG	Reference
SAP6	AP6	Common bacterial blight (CBB)	820 cis	F - GTC ACG TCT CCT TAA TAG TA R - GTC ACG TCT CAA TAG GCA AA	Major QTL (GN#1 sel 27)	B10	Miklas et al., 2000b,c
BAC6	BC409	CBB	1250 cis	TAG GCG GCG GCG CAC GTT TTG TAG GCG GCG GAA GTG GCG GTG	Major QTL (GN#1 sel 27)	B10	Jung et al., 1999
*S*U91	U9	CBB	700 cis	CCA CAT CGG TTA ACA TGA GT CCA CAT CGG TGT CAA CGT GA	Major QTL (XAN 159)	B8	Pedraza et al., 1997
LG5 syn. BC420	BC420	CBB	900 cis	GCA GGG TTC GAA GAC ACA CTG G GCA GGG TTC GCC CAA TAA CG	Major QTL (XAN 159)	B6	Yu et al., 2000
R7313		CBB	700 cis	ATT GTT ATC GTC GAC ACG AAT ATT TCT GAT CAC ACG AG	Major QTL (OAC 88-1)	B8	Bai et al., 1997 Beattie et al., 1998
R4865		CBB	950 cis	TCC AAA GCC ATT CTA GTT CAG CTA CTT TCA AAC TGG G	Major QTL (OAC 88-1)		Bai et al., 1997 Beattie et al., 1998
SB10	B10	Halo bacterial blight (HBB)	525 cis	CTG CTG GGA CAA TCA CCA AGT C CTG CTG GGA CTC TCT TAC	*Pse-1*	B4	Fourie et al., 2004
*S*W13	W13	Bean common mosaic virus (BCMV) & HBB	690 cis	CAC AGC GAC ATT AAT TTT CCT TTC CAC AGC GAC	/ *Pse-3*	B2	Haley et al., 1994 Melotto et al., 1996 Fourie et al.,

				AGG AGG AGC TTA TTA			2004
ROC11	C11	BCMV	420 trans	CCA ATT CTC TTT CAC TTG TAA CC GCA TGT TCC AGC AAA CC	*bc-3*	B6	Johnson et al., 1997
*S*BD5	BD5	BCMV	1250 cis	GTG CGG AGA GGC CAT CCA TTG GTG GTG CGG AGA GTT TCA GTG TTG ACA	*bc-1*^{2}	B3	Miklas et al., 2000a
*S*R2	R2	Bean golden yellow mosaic virus (BGYMV)	530 / 570 codominant	CAC AGC TGC CCT AAC AAA AT CAC AGC TGC CAC AGG TGG GA	*bgm-1*		Urrea et al., 1996 Beebe, PC, 1996
*S*W12	W12	BGYMV	700 cis	TGG GCA GAA GTT CTA GCA TGT GGC TGG GCA GAA GCA CAG TAT GAT TTG	Major QTL (DOR 364)	B4	Miklas et al., 2000c Singh et al. 2000
SAS8	AS08	Beet curly top virus (BCTV)	1550 cis	GGC TGC CAG TAT CTT GTC TAA CAC C GGC TGC CAG TGA CGC AAT TCT GCA G	*Bct*	B7	Larsen and Miklas, 2004
*S*K14	K14	Rust	620 cis	CCC GCT ACA CAC CAA TAC CTG CCC GCT ACA CTT GAT AAA ATG TTA G	*Ur-3*	B11	Haley et al. 1994 Nemchin. & Stavely, 1998 Miklas et al., 2002

SA14	A14	Rust	1079 / 800 codominant	CTA TCT GCC ATT ATC AAC TCA AAC GTG CTG GGA AAC ATT ACC TAT T	*Ur-4*	B6	Miklas et al. 1993 Miene et al. 2004 Miklas et al., 2002
*SI*19	I19	Rust	460 cis	AAT GCG GGA GAT ATT AAA AGG AAA G AAT GCG GGA GTT CAA TAG AAA AAC C	*Ur-5*	B4	Haley et al., 1993 Melotto et al., 1998 Miklas et al., 2000c
SBC6	BC06	Rust	308 cis	GAA GGC GAG AAG AAA AAG AAA AAT GAA GGC GAG AGC ACC TAG CTG AAG	*Ur-6*	B11	Park et al., 2003b, 2004b Miklas et al. 2002
SAD12	AD12	Rust	537 cis	AAG AGG GCG TGA GAT CGT CG AAG AGG GCG TCT TGA AGG TT	*Ur-7*	B11	Park et al., 2003a, 2004a
SAE19	AE19	Rust	890 trans	CAG TCC CTG ACA ACA TAA CAC C CAG TCC CTA AAG TAG TTT GTC CCT A	*Ur-11*	B11	Johnson et al., 1995 Alzate-Marin et al., 2004 Queiroz et al., 2004c Miklas et al., 2002
UR11-GT2	GT02	Rust	450 cis	CGC ACT TAG GAG CAC AAA TGG TGG GTC CCA TAT TTT G	*Ur-11*	B11	Boone et al., 1999 *Miklas et al., 2002
KB126	E-AAC/M-ACC	Rust	405 / 430 codominant	GAA TTC AAC CTC GGC CAC TAC C TTA AAC CTT CCG GAG GAT TC	*Ur-13*	B8	Mienie et al., 2005
SF10	F10	Rust	1072 cis	GGA AGC TTG GTG AGC AAG GA GGA AGC TTG GCT ATGATG GT	Ouro Negro	B4	Correa et al., 2000 Faleiro et al. 2000a Miklas et al.,

							2002
SBA8	BA8	Rust	530 cis	CCA CAG CCG ACG GAG GAG GCC ATG TTT TTT GTC CCC	Ouro Negro	B4	Correa et al., 2000 Miklas et al., 2002
Phs	Phaseolin 'T' & 'S' alleles	White mold & CBB	Multiple	AGC ATA TTC TAG AGG CCT CC GCT CAG TTC CTC AAT CTG TTC	Major QTL (G 122) & (BAT 93)	B7	Kami et al., 1995 Nodari et al., 1993 Miklas et al., 2001
SAU5	AU05	White mold	1350 cis	GAG CTA CCG TCA GTT TAC TAA GAG CTA CCG TGG CTT TTT TCT	QTL (minor) NY6020-4	B6	Miklas et al., 2003
SS18	S18	White mold	1650 cis	CTG GCG AAC TGT ACA TGC AAC ATA C CTG GCG AAC TGA TTC ATA CAT TTT G	QTL (major) NY6020-4	B8	Miklas et al., 2003
	SE_{ACT}/M_{CCA}	Anthracnose	codominant		*Co-1*[2]	B1	Vallejo and Kelly, 2002
SCAreoli	H20	Anthracnose	1000	GGG AGA CAT CCA TCA GAC AAC TCC GGG AGA CAT CTT CAT TTG ATA TGC	*Co-2*	B11	Geffroy et al., 1998 Adam-Blondon et al., 1994
SY20	Y20	Anthracnose	830 cis	AGC CGT GGA AGG TTG TCA T CCG TGG AAA CAA CAC ACA AT	*Co-4*	B8	Arruda et al. 2000 Queiroz et al., 2004b *Kelly et al., 2003

SC08	C08	Anthracnose	910 cis	AGA ATG CCT TTA GCT GTT GG CAG AGA GGC TAG GCT TAT CG	*Co-4*	B8*	Arruda et al. 2000 Queiroz et al., 2004b Kelly et al., 2003
*S*AS13	AS13	Anthracnose	950 cis	CAC GGA CCG AAT AAG CCA CCA ACA CAC GGA CCG AGG ATA CAG TGA AAG	*Co-4*^{2}	B8	Young et al., 1998 Kelly et al., 2003
SH18	H18	Anthracnose	1100 cis	CCA GAA GGA GCT GAT AGT ACT CCA CAA C GGT AGG CAC ACT GAT GAA TCT CAT GTT GGG	*Co-4*^{2}	B8	Awale and Kelly, 2001 Alzate-Marin et al., 2001 Kelly et al., 2003
SBB14	BB14	Anthracnose	1150/1050 codominant	GTG GGA CCT GTT CAA GAA TAA TAC GTG GGA CCT GGG TAG TGT AGA AAT	*Co-4*^{2}	B8	Awale and Kelly, 2001 Kelly et al., 2003
SAB3	AB-3	Anthracnose	400 cis	TGG CGC ACA CAT AAG TTC TCA CGG TGG CGC ACA CCA TCA AAA AAG GTT	*Co-5*		Vallejo and Kelly, 2001
SAZ20	AZ20	Anthracnose	845 cis	ACC CCT CAT GCA GGT TTT TA CAT AAT CCA TTC ATG CTC ACC	*Co-6*	B7	Alzate-Marin et al., 2000 Queiroz et al., 2004b *Kelly et al., 2003
SZ04	Z04	Anthracnose	567 cis	GGC TGT GCT GAT TAA TTC TGG TGC TCA TTT TAT AAT GGA GAA AAA	*Co-6*	B7	Alzate-Marin et al., 1999 Queiroz et al., 2004b Kelly et al., 2003

SB12	B-12	Anthracnose	350 cis	CCT TGA CGC ACC TCC ATG TTG ACG ATGGG TTG GCC	*Co-9*	B4	Mendez de Vigo et al., 2002
SF10	F10	Anthracnose	1072 cis	GGA AGC TTG GTG AGC AAG GA GGA AGC TTG GCT ATGATG GT	*Co-10*	B4	Correa et al., 2000 Faleiro et al., 2000b Alzate-Marin et al., 2003
SH13	H13	Angular leaf spot	520 cis	GAC GCC ACA CCC ATT ATG TT GCC ACA CAG ATG GGA GCT TTA	AND 277 dominant gene *Phg-1*		Carvalho et al. (1998) Queiroz et al., 2004a
SN02	N02	Angular leaf spot	890 cis	ACC AGG GGC ATT ATG AAC AG ACC AGG GGC AAC ATA CTA TG	Mexico 54 & C49-242 dominant gene *Phg-2*	B8	Sartorato et al., 2000 Nietsche et al., 2000 Miklas, PC, 2002
	E-ACA/M-CTT_{330}	Angular leaf spot	280 / 305 codominant	CTT GTT CTG AGT CAT TTA CCT TGC GAA TTC ACA GTC CAA ACT CTA ATC	G 10474 Dominant gene		Mahuku et al., 2004
SAA19	AA19	Angular leaf spot	650 cis	TGA GGC GTG TCA ATG GAT ATA A GAG GCG TGT TGA TAA TTC TGG	Ouro Negro dominant gene		Corrêa et al. 2001 Queiroz et al., 2004a
SBA16	BA16	Angular leaf spot	560 cis	TTC CAC GTC TAT TTT GCA TCA CAC GCA TCA CGC AGA ACT	Ouro Negro dominant gene		Faleiro et al., 2003 Queiroz et al., 2004a
SM02	M02	Angular leaf spot	460 cis	CAA CGC CTC ATT AAA TTG GA CGC CTC TAA ACG GGA GAA AC	Ouro Negro dominant gene		Corrêa et al., 2001 Queiroz et al., 2004a

SCAR Name	**PCR Protocol**
*S*AP6	34 cycles of 10s at 94°C, 40s at 55°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
*S*U91	34 cycles of 10s at 94°C, 40s at 58°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
LG5	35 cycles of 30s at 94°C, 30s at 50°C, and 60s at 72°C; followed by one cycle of 5 minutes at 72°C
R7313	34 cycles of 10s at 94°C, 40s at 60°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
R4865	34 cycles of 10s at 94°C, 40s at 60°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
BC409	34 cycles of 10s at 94°C, 60s at 70°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
*S*W13	34 cycles of 10s at 94°C, 40s at 67°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
ROC11	34 cycles of 10s at 94°C, 40s at 55°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
*S*BD5	34 cycles of 10s at 94°C, 40s at 65°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
*S*R2	34 cycles of 10s at 94°C, 40s at 60°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C 60°C annealing = codominant; 65°C annealing = dominant
*S*W12	1 cycle of 60s at 94°C; 30 cycles of 30s at 94°C, 30s at 70°C, and 60s at 72°C; followed by one cycle of 5 minutes at 72°C
SAS8	1 cycle of 5 min at 94°C; 30 cycles of 60s at 94°C, 60s at 68°C, and 60s at 72°C; followed by one cycle of 7 minutes at 72°C

*S*K14	34 cycles of 10s at 94°C, 40s at 63°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
*S*I19	34 cycles of 10s at 94°C, 40s at 67°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
Ur-6	94°C 2 min; 30 cycles of 30s at 94°C, 60s at 59°C, and 120s at 72°C; followed by one cycle of 7 minutes at 72°C
UR11-GT2	60°C annealing = codominant; 65°C annealing = dominant
SF10	1 cycle of 94°C of 3 min: 35 Cycles at 15s at 94°C, 60s at 65°C and 90s at 72°C; followed by 1 cycle of 7 min at 72°C
SBA8	1 cycle of 94°C of 3 min; 35 Cycles at 15s at 94°C, 60s at 65°C and 90s at 72°C; followed by 1 cycle of 7 min at 72°C
Phs	34 cycles of 10s at 94°C, 40s at 50°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
SAU5	34 cycles of 10s at 94°C, 40s at 60°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
SS18	34 cycles of 10s at 94°C, 40s at 63°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
SCAreoli	58°C annealing temperature followed by *Dra*I digestion
S*A*S13	34 cycles of 10s at 94°C, 144s at 72°C; followed by one cycle of 5 minutes at 72°C
SH18	34 cycles of 10s at 94°C, 40s at 62°C, and 120s at 72°C; followed by one cycle of 5 minutes at 72°C
SBB14	34 cycles of 10s at 94°C, 40s at 62°C, and 120s at 72°C: followed by one cycle of 5 minutes at 72°C
SAB3	1 cycle of 3 minutes at 94°C; 30 cycles of 10s at 94°C, 30s at 65°C, 2 minutes at 72°C; followed by 1 cycle of 5 minutes at 72°C
SB12	1 cycle of 94°C of 2 min: 35 Cycles of 60s at 94°C, 60s at 68°C, and 60s at 72°C; followed by 1 cycle of 7 min at 72°C
SN02	30 Cycles of 30s at 94°C, 60s at 65°, and 90s at 72°C

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